

# **DNA Lesion Can Facilitate Base Ionization: Vertical Ionization Energies of Aqueous 8-Oxoguanine and its Nucleoside and Nucleotide**

*Vladimír Palivec,<sup>1</sup> Eva Pluhařová,<sup>1+</sup> Isaak Unger,<sup>2</sup> Bernd Winter,<sup>2</sup> and Pavel Jungwirth<sup>1\*</sup>*

<sup>1</sup>Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, Flemingovo nám. 2, 16610 Prague 6, Czech Republic

<sup>2</sup>Institute of Methods for Material Development, Helmholtz Center Berlin, Albert-Einstein-Strasse 15, D-12489 Berlin, Germany

<sup>+</sup>*Present address: Department of Chemistry, École Normale Supérieure, UMR ENS-CNRS-UPMC 8640, 24 rue Lhomond, 75005 Paris, France*

<sup>\*</sup>*Corresponding author: [pavel.jungwirth@uochb.cas.cz](mailto:pavel.jungwirth@uochb.cas.cz)*

## ABSTRACT

8-oxoguanine is one of the key products of indirect radiation damage to DNA by reactive oxygen species. Here, we describe ionization of this damaged nucleobase and the corresponding nucleoside and nucleotide in aqueous phase, modeled by the non-equilibrium polarizable continuum model, establishing their lowest vertical ionization energies of 6.8 - 7.0 eV. We thus confirm that 8-oxoguanine has even lower ionization energy than the parental guanine, which is the canonical nucleobase with the lowest ionization energy. Therefore, it can act as a trap for the cationic hole formed by ionizing radiation and thus protect DNA from further radiation damage. We also model using time-dependent density functional theory and measure by liquid jet photoelectron spectroscopy the valence photoelectron spectrum of 8-oxoguanine in water. We show that the calculated higher lying ionization states match well the experiment which, however, is not sensitive enough to capture the electron signal corresponding to the lowest ionization process due to the low solubility of 8-oxoguanine in water.

**KEYWORDS:** 8-oxoguanine, aqueous solution, photoionization, ab initio calculations

## Introduction

Ionization of nucleobases is the initial key step leading to direct DNA damage and mutation.<sup>1</sup> Among the four nucleobases guanine has the lowest ionization energy.<sup>2</sup> Therefore, the positive hole created in the photoionization process tends to localize on the guanine base, which is also the most susceptible site for oxidative processes. Vulnerability of guanine to reactive oxygen species formed among others within indirect radiation damage of DNA leads to production of a variety of products, in particular 8-oxoguanine (8-OG).<sup>3</sup> This lesion is estimated to be generated at the rate of approximately 2000 per human cell per day.<sup>4,5</sup> Its concentration in the cellular DNA is, in fact, a quantitative measure of the degree of damage that an organism has undergone.<sup>6</sup> The 8-OG lesions within DNA can cause serious problems.<sup>7,8</sup> Because of an additional oxygen atom on the base it can form a Hoogsten base pair with adenine instead of forming the common guanine-cytosine Watson-Crick base pair. It is believed that this base pair mismatch is responsible for incorrect interpretation of genetic code, and consequently may lead to mutations.<sup>9,10</sup> On the other hand, 8-OG is more susceptible to oxidation than the original base guanine. Because of this, it has been suggested that free 8-OG may serve as the trap for the positive hole and thus protect other bases including guanine from oxidation.<sup>8,11</sup>

Further oxidation of 8-OG has been studied both by experimental<sup>12,13</sup> and theoretical<sup>14</sup> groups. Additionally, there are experimental<sup>11</sup> and computational<sup>15</sup> studies of ionization, which may precede oxidation of guanine to 8-OG, however, the crucial knowledge of the vertical ionization energy (VIE) of 8-OG and its nucleoside and nucleotide in the context of the aqueous DNA environment has been missing, with only a single calculation on the parent species published most recently.<sup>16</sup> The principle aim of the present study is to fill this gap by establishing computationally accurate VIEs of the aqueous 8-OG base and its nucleoside and nucleotide.

## Methods

### *Computational*

The lowest VIEs of 8-OG and its singly ( $8\text{-OG}^{1-}$ ) and doubly ( $8\text{-OG}^{2-}$ ) deprotonated forms, as well as 8-oxo-2'-deoxyguanosine (8-OdGs) and the singly deprotonated 8-oxo-2'-deoxyguanosine monophosphate ( $8\text{-OdGMP}^{1-}$ ) were evaluated as the difference between the ground state energy after and before ionization, at the optimal geometry of the closed shell species before ionization. For calculating higher VIEs for 8-OG,  $8\text{-OG}^{1-}$ , and  $8\text{-OG}^{2-}$  excitation energies to the singly occupied molecular orbital of the ionized molecule (again at the geometry of the species before ionization) were added to the lowest VIE.<sup>17, 18</sup> By Gaussian broadening each of the calculated ionization energy by 1 eV<sup>17, 18</sup> this procedure allowed us to construct photoionization spectra comparable to experiment. For calculation of the spectra, we considered only the lowest in energy tautomer of  $8\text{-OG}^{1-}$  and  $8\text{-OG}^{2-}$ , which should be dominantly populated (~90 %) at ambient conditions.<sup>19</sup> Additionally, test calculations for the second lowest tautomers showed that the spectra are very similar to those for the lowest tautomers.

Effects of solvation were taken into account within the non-equilibrium polarizable continuum model (NEPCM) accounting for the electronic but not nuclear relaxation of the solvent upon ionization of the solute, which is appropriate when modeling VIEs.<sup>20-22</sup> For the doubly charged  $8\text{-OG}^{2-}$  species we checked the applicability of the continuum description of the solvent by performing also hybrid calculations with one to five explicit water molecules optimized around  $8\text{-OG}^{2-}$  and then placed in the NEPCM cavity. Within this procedure, excitations originating from the explicit water molecules were excluded from the spectra and only excitations originating from the base were considered.

To evaluate the lowest VIE, we used the unrestricted version of the second order Møller-Plesset (MP2) method with the aug-cc-pVDZ basis set. Higher spin components were annihilated via Schlegel's projection method (PMP2).<sup>23</sup> Previous calculations for isolated gas-phase nucleic acid bases showed that for purine nucleobases this approach provides ionization energies within 0.2 eV from the benchmark CCSD(T) values.<sup>24</sup> To obtain the higher ionization states, we additionally employed the time-dependent density functional approach (TDDFT) using the BMK functional,<sup>25</sup> as in our previous study on purines.<sup>17</sup> All calculations were performed using the Gaussian 03 program.<sup>26</sup>

### *Experimental*

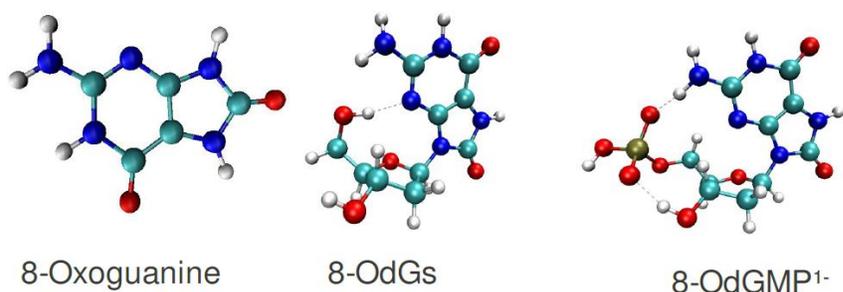
Valence photoelectron-spectroscopy measurements from 8-OG (2-amino-6,8-dihydroxypurine hydrochloride) aqueous solution were done at the U41-PGM undulator beamline of the synchrotron radiation facility BESSY II, Berlin. All the spectra were collected from a 20- $\mu\text{m}$  vacuum liquid jet travelling at a velocity of approximately  $40 \text{ ms}^{-1}$ , with a temperature of  $10^\circ\text{C}$ . Details of the technique and experimental setup were described in our previous papers.<sup>27, 28</sup> In short, photoelectrons were detected in the direction normal to both the synchrotron light polarization vector and the flow of the liquid jet. Photoelectrons pass through a 150  $\mu\text{m}$  diameter orifice separating the main interaction chamber (at  $10^{-4}$  mbar) from the differentially pumped detector chamber (at  $10^{-8}$  mbar), which houses a hemispherical electron-energy analyzer. The distance of less than 0.5 mm between the liquid jet and the orifice ensures that the detected electrons did not suffer from inelastic scattering with water vapor near the jet surface.<sup>27</sup> The applied photon energy was 180 eV and the energy resolution of the beamline in this energy range was better than 50 meV. The resolution of the hemispherical energy analyzer of approximately 100 meV at 10 eV pass energy is constant with kinetic energy. The small focal size of 23x12

$\mu\text{m}^2$  of the incident photon beam matches the diameter of the liquid jet leading to an almost negligible photoelectron signal from water vapor (less than 5% of the total signal).

In order to achieve the highest possible concentration, 8-OG has been dissolved in water at basic pH. We found that at pH 12.6 500 mg of 8-OG is fully soluble up to 0.04 M, but this is not the case at lower pH. Adjustment of pH was made by addition of NaOH. The basic water solution, containing no 8-OG, was used to measure a reference photoelectron spectrum, and its subtraction from the solution spectrum yields the small signal due to ionization of 8-OG(aq). 2-amino-6,8-dihydropurine hydrochloride (purity >90%) was purchased from Toronto Research Chemicals Inc., and was used here without further purification. A consequence of the necessity to use a highly alkaline solution to increase solubility of 8-OG is that it becomes deprotonated. Indeed, since the pKas of the N1 and N9 sites are estimates as 8.7 and 11.9, respectively,<sup>19</sup> in our experiment 8-OG is in a mixture of about 83 % of 8-OG<sup>2-</sup> and about 17 % of 8-OG<sup>1-</sup>.

## Results and Discussion

Figure 1 shows the most stable aqueous structures of 8-OG, 8-OdGs, and 8-OdGMP<sup>1-</sup>. Note that both mono- and di-anionic nucleotides are present in the solution at neutral pH since the corresponding pKa<sub>2</sub> of phosphate is 7.2.<sup>29</sup> However, only the former is relevant in the DNA context. Table 1 presents the corresponding vertical ionization energies (VIE) calculated using the non-equilibrium polarizable continuum (NEPCM)<sup>20-22</sup> to model the aqueous environment (for details see Computational Methods). For reference we show also our previously calculated results for guanine and its derivatives<sup>17</sup> and compare results in water with those in the gas phase. In water, the lowest ionization always originates from the base, as demonstrated in Figure 2 which depicts the highest occupied molecular orbital (HOMO) of 8-OdGMP<sup>1-</sup>.

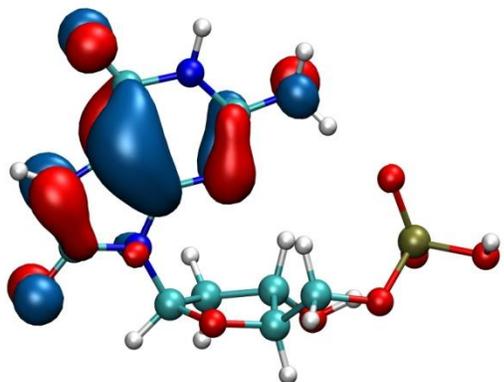


**Figure 1:** Most stable aqueous-phase structures of 8-oxoguanine and its nucleoside and nucleotide.

**Table 1.** Vertical ionization energies (VIE) in eV calculated at the PMP2/aug-cc-pVDZ level in the aqueous phase employing NEPCM and in the gas phase.

		nucleobase	deoxynucleoside	deoxynucleotide <sup>1-</sup>
Aqueous phase	8-Oxoguanine	6.94	7.01	6.79
	Guanine	7.34	7.42 <sup>a</sup>	7.08 <sup>a</sup>
Gas phase	8-Oxoguanine	7.84	7.98	5.09
	Guanine	8.43	8.38 <sup>a</sup>	5.18 <sup>a</sup>

<sup>a</sup>VIE of the ribose analogues from Ref. <sup>17</sup>, with that the difference between VIEs of deoxyribose vs. ribose containing nucleosides and nucleotides is negligible.



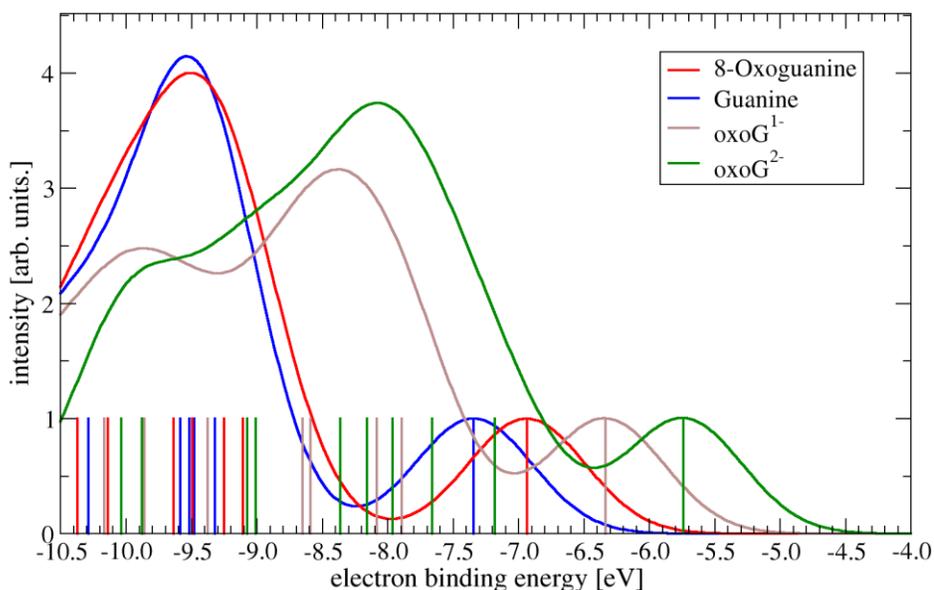
**Figure 2:** The highest occupied molecular orbital (HOMO) of aqueous 8-OdGMP<sup>1-</sup> demonstrating that the lowest ionization originates from the base.

Similarly to the canonical bases,<sup>17, 18</sup> we see from Table 1 the remarkable ability of the aqueous environment to screen the effect of the sugar and, in particular, of the phosphate on the ionization energy of 8-OG. While in the gas phase the phosphate anionic group strongly destabilizes the base leading to lowering of the VIE by almost 3 eV, in water this effect practically vanishes (Table 1). Most importantly, Table 1 demonstrates that in water 8-OG has VIE lower by about 0.4 eV than guanine, and this behavior is semi-quantitatively preserved also for the corresponding aqueous nucleosides and nucleotides.<sup>17</sup> Qualitatively, the reason for the lowering of VIE compared to guanine is the presence of the electronegative oxygen atom in 8-OG, which provides additional electron density and thus destabilizes the part of the molecules from which ionization originates. We also mention that the present value of VIE for 8-OG is very close to the value of 7.1 eV calculated most recently for the same species using a similar approach.<sup>16</sup> Finally, we have shown previously that in water the DNA environment has little effect on the VIE.<sup>30</sup> Our calculations thus support the hypothesis that not only free 8-OG but potentially also that produced in DNA could serve as a protective sink of the cationic hole upon prolonged exposure of DNA to ionizing radiation.<sup>11</sup> At the same time, lowering of the ionization

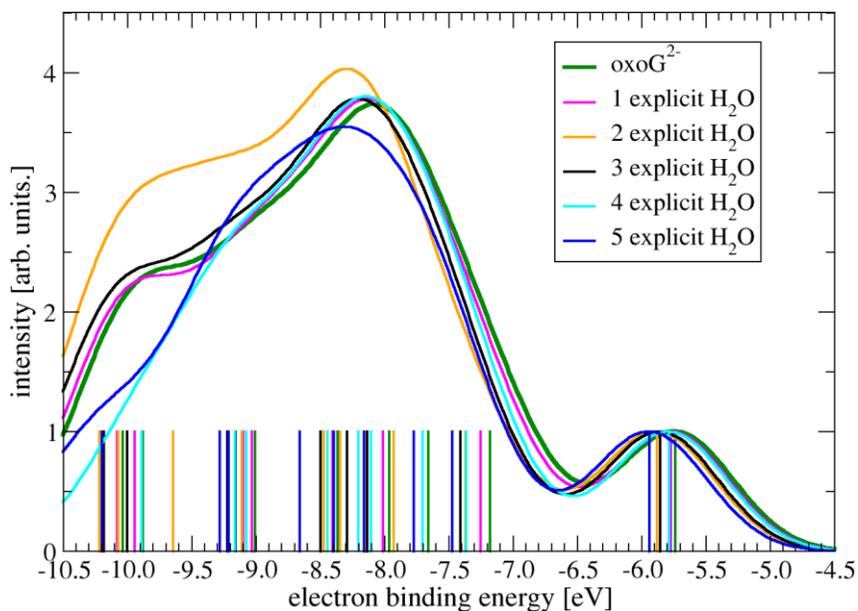
energy due to the presence of 8-OG increases the chance of moving the photoionization threshold of damaged DNA toward the edge of the terrestrial solar UV spectrum, as has already been suggested for undamaged DNA.<sup>31</sup>

In order to facilitate direct comparison with experiment (*vide infra*) we evaluated in addition to the lowest VIE also higher ionized states of 8-OG, 8-OG<sup>1-</sup>, and 8-OG<sup>2-</sup>. Each of these calculated peaks was then broadened with a Gaussian with a full-width-at-half-maximum (FWHM) of 1 eV to account for the experimental peak broadening assuming the same cross section for all transitions.<sup>18, 32</sup> The resulting spectra are presented in Figure 3 and compared to that of the aqueous canonical guanine base. We see that oxidation causes a shift to lower energies not only for the lowest ionization but also for the higher-lying ionized states. However, the shift for the lowest state is the largest, while that for the higher state gets to a large extent buried within the width of the higher energy peak (Figure 3). Furthermore, deprotonation directly on the base further shifts the ionization energies to even lower values. We mention in passing that this is different from the situation when deprotonation occurs on the phosphate moiety of the corresponding nucleotide, which has only a very small effect on the position of the lowest ionization peak.<sup>17, 18</sup>

For the singly and, in particular, the doubly deprotonated species, one may question the validity of the continuum approach to solvation due to potential strong perturbation of the solvent by the charged solute. In order to test this effect, we performed additional spectral calculations for 8-OG<sup>2-</sup> including one to five explicit water molecules into the NEPCM cavity. The resulting spectra are presented in Figure 4. We see that in overall the continuum model performs surprisingly well even for 8-OG<sup>2-</sup> with the lowest energy peak being practically unaltered and the higher peaks only moderately modified by including explicit water molecules.

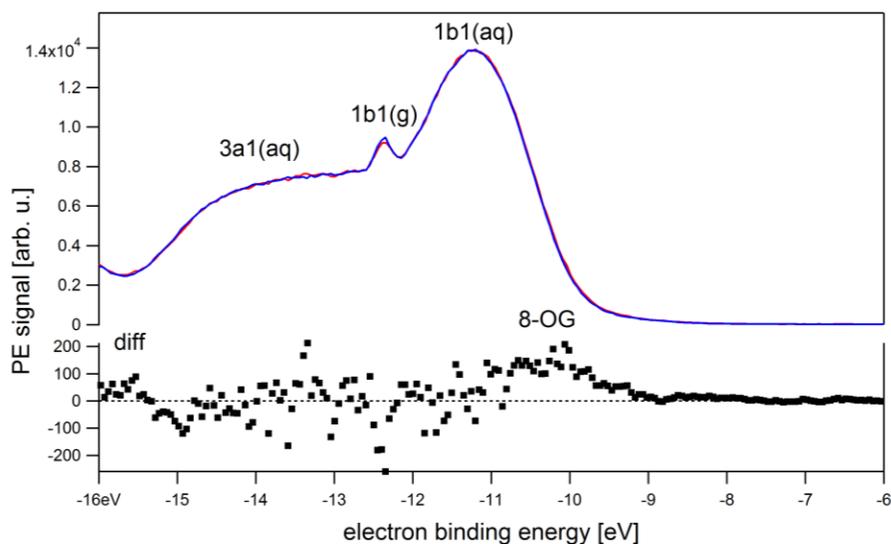


**Figure 3:** Photoionization spectrum of aqueous 8-oxoguanine (red) compared to that of the canonical guanine base (blue) in water. In addition, spectra of the singly and doubly deprotonated 8-oxoguanine are depicted in brown and green.



**Figure 4:** Photoionization spectrum of aqueous doubly deprotonated 8-oxoguanine,  $8\text{-OG}^{2-}$ , with 0 – 5 explicit water molecules included into the NEPCM solvent cavity.

We also attempted to obtain a photoelectron (PE) spectrum of 8-OG aqueous solution experimentally (for details see Experimental Methods) in order to benchmark the calculations. Due to the high pH of 12.6 employed in the experiment to increase solubility, the investigated species is actually a mixture of doubly ( $8\text{-OG}^{2-}$ ) and singly ( $8\text{-OG}^{1-}$ ) deprotonated forms, denoted further as  $8\text{-OG}^{1-/2-}$ . Still, because of the very low solubility in water even at higher pH, signal-to-noise in our spectra allows to unequivocally identify only states with energies larger than the lowest ionization energy, which have larger integrated ionization probability. Results are shown in Figure 5 presenting the PE spectrum from the 0.04 M  $8\text{-OG}^{1-/2-}$  basic aqueous solution (in red) along with the water reference (in blue) spectrum, both measured under identical experimental conditions at a photon ionization energy of 180 eV. Despite using high pH, the signal corresponding to the lowest ionization energy of  $8\text{-OG}^{1-/2-}(\text{aq})$  is within the experimental noise level, and cannot thus be quantified here. Yet, there seems to be an indication of small above-zero signal from the base in the differential PE spectrum (in black) near 8.2 eV electron binding energy, which correlates well with the second ionization peak of the calculated spectrum of both  $8\text{-OG}^{1-}$  and  $8\text{-OG}^{2-}$ . This signal is observable here due to the larger integrated ionization probability than for lowest ionization energy. The experimental signal intensity in the differential spectrum near 9-10.5 eV electron binding energy, which has been successfully used in previous studies to single out solute contributions,<sup>32,33</sup> is in agreement in position, albeit stronger in intensity, compared to the calculated additional peak arising from the higher VIEs for both  $8\text{-OG}^{1-}$  and  $8\text{-OG}^{2-}$  (Figure 3). Note that analysis of the experimental spectrum for binding energies larger  $\sim 10.5$  eV is not feasible because the water signal fully overwhelms that from  $8\text{-OG}^{1-/2-}$  and, consequently, the differential spectrum only reports noise.



**Figure 5:** Photoelectron spectrum from 0.04 M 8-oxoguanine (i.e., mixture of its singly and doubly deprotonated forms  $8\text{-OG}^{1-2-}$ ), aqueous solution (red) and of a reference spectrum from liquid water at pH 12.6 (blue), both measured at 180 eV photon energy. The differential spectrum, solution minus water signal, is shown at the bottom (in black squares). Top labels indicate the water valence orbitals which are ionized. The small narrow peak,  $1b_1(g)$ , arises from ionization of gas-phase water's highest occupied molecular orbital.

## Conclusions

In summary, using quantum chemical calculations we have shown that 8-oxoguanine, which is a primary product of radiation damage to DNA, has in the aqueous environment ionization potential lower by about 0.4 eV compared to the canonical guanine base. The calculated photoelectron spectra agree semi-quantitatively with results from liquid microjet photoelectron spectroscopy experiments where, however, only the higher ionization states between 8 and 10.5 eV of can be discerned due to the low solubility of 8-oxoguanine in water, leading to small electron signal intensity. Moreover, the necessity to use high pH conditions in

the experiment to increase solubility of the base results in formation of a mixture of singly and doubly deprotonated 8-oxoguanine. The present results strongly support the hypothesis<sup>11</sup> that due to its low ionization potential 8-oxoguanine can serve as a sink to the cationic hole and, therefore, help to protect DNA from further radiation damage.

## **Acknowledgment**

Support from the Czech Science Foundation (grant P208/12/G016) and the Academy of Sciences (Praemium Academie award) is gratefully acknowledged. EP thanks the IMPRS Dresden for support. Allocation of computer time from the National supercomputing center IT4Innovations in Ostrava is appreciated.

## **References**

- (1) Hall, D. B.; Holmlin, R. E.; Barton, J. K. Oxidative DNA Damage through Long-Range Electron Transfer. *Nature* **1996**, *382*, 731-735.
- (2) Ward, J. F. Nature of Lesions Formed by Ionizing Radiation. In *DNA Damage and Repair*, Nickoloff, J. A.; Hoekstra, M. F., Eds. Humana Press: Totowa, N.J., 1998; Vol. 2, pp 65-84.
- (3) Cullis, P. M.; Malone, M. E.; MersonDavies, L. A. Guanine Radical Cations are Precursors of 7,8-dihydro-8-oxo-2'-deoxyguanosine but are not Precursors of Immediate Strand Breaks in DNA. *J. Am. Chem. Soc.* **1996**, *118*, 2775-2781.
- (4) Beckman, K. B.; Ames, B. N. Oxidative Decay of DNA. *J. Biol. Chem.* **1997**, *272*, 19633-19636.
- (5) Foksinski, M.; Rozalski, R.; Guz, J.; Ruszkowska, B.; Sztukowska, P.; Piwowarski, M.; Klungland, A.; Olinski, R. Urinary Excretion of DNA Repair Products Correlates with Metabolic Rates as well as with Maximum Life Spans of Different Mammalian Species. *Free Radical Biol. Med.* **2004**, *37*, 1449-1454.

- (6) Kasai, H. Analysis of a Form of Oxidative DNA Damage, 8-hydroxy-2'-deoxyguanosine, as a Marker of Cellular Oxidative Stress During Carcinogenesis. *Mutation Research-Rev. Mutation Res.* **1997**, *387*, 147-163.
- (7) Koizume, S.; Inoue, H.; Kamiya, H.; Ohtsuka, E. Neighboring Base Damage Induced by Permanganate Oxidation of 8-oxoguanine in DNA. *Nucleic Acids Res.* **1998**, *26*, 3599-3607.
- (8) Kim, J. E.; Choi, S.; Yoo, J. A.; Chung, M. H. 8-oxoguanine Induces Intramolecular DNA Damage but Free 8-oxoguanine Protects Intermolecular DNA from Oxidative Stress. *Febs Lett.* **2004**, *556*, 104-110.
- (9) Neeley, W. L.; Essigmann, J. M. Mechanisms of Formation, Genotoxicity, and Mutation of Guanine Oxidation Products. *Chem. Res. Toxicol.* **2006**, *19*, 491-505.
- (10) Bruner, S. D.; Norman, D. P. G.; Verdine, G. L. Structural Basis for Recognition and Repair of the Endogenous Mutagen 8-oxoguanine in DNA. *Nature* **2000**, *403*, 859-866.
- (11) Steenken, S.; Jovanovic, S. V.; Bietti, M.; Bernhard, K. The Trap Depth (in DNA) of 8-oxo-7,8-dihydro-2'-deoxyguanosine as Derived from Electron-Transfer Equilibria in Aqueous Solution. *J. Am. Chem. Soc.* **2000**, *122*, 2373-2374.
- (12) Niles, J. C.; Wishnok, J. S.; Tannenbaum, S. R. Spiroiminodihydantoin and Guanidinohydantoin are the Dominant Products of 8-oxoguanosine Oxidation at Low Fluxes of Peroxynitrite: Mechanistic Studies with <sup>18</sup>O. *Chem. Res. Toxicol.* **2004**, *17*, 1510-1519.
- (13) Yu, H. B.; Venkatarangan, L.; Wishnok, J. S.; Tannenbaum, S. R. Quantitation of Four Guanine Oxidation Products from Reaction of DNA with Varying Doses of Peroxynitrite. *Chem. Res. Toxicol.* **2005**, *18*, 1849-1857.

- (14) Munk, B. H.; Burrows, C. J.; Schlegel, H. B. An Exploration of Mechanisms for the Transformation of 8-oxoguanine to Guanidinohydantoin and Spiroiminodihydantoin by Density Functional Theory. *J. Am. Chem. Soc.* **2008**, *130*, 5245-5256.
- (15) Psciuk, B. T.; Lord, R. L.; Munk, B. H.; Schlegel, H. B. Theoretical Determination of One-Electron Oxidation Potentials for Nucleic Acid Bases. *J. Chem. Theo. Comput.* **2012**, *8*, 5107-5123.
- (16) Sieradzan, I.; Marchaj, M.; Anusiewicz, I.; Skurski, P.; Simons, J. Prediction of Thymine Dimer Repair by Electron Transfer from Photoexcited 8-Aminoguanine or Its Deprotonated Anion. *J. Phys. Chem. A* **2014**, *118*, 7194-7200.
- (17) Pluharova, E.; Jungwirth, P.; Bradforth, S. E.; Slavicek, P. Ionization of Purine Tautomers in Nucleobases, Nucleosides, and Nucleotides: From the Gas Phase to the Aqueous Environment. *J. Phys. Chem. B* **2011**, *115*, 1294-1305.
- (18) Slavicek, P.; Winter, B.; Faubel, M.; Bradforth, S. E.; Jungwirth, P. Ionization Energies of Aqueous Nucleic Acids: Photoelectron Spectroscopy of Pyrimidine Nucleosides and ab Initio Calculations. *J. Am. Chem. Soc.* **2009**, *131*, 6460-6467.
- (19) Jang, Y. H.; Goddard, W. A.; Noyes, K. T.; Sowers, L. C.; Hwang, S.; Chung, D. S. First Principles Calculations of the Tautomers and pK<sub>a</sub> Values of 8-oxoguanine: Implications for Mutagenicity and Repair. *Chem. Res. Toxicol.* **2002**, *15*, 1023-1035.
- (20) Cossi, M.; Barone, V. Separation between Fast and Slow Polarizations in Continuum Solvation Models. *J. Phys. Chem. A* **2000**, *104*, 10614-10622.
- (21) Cossi, M.; Barone, V. Solvent Effect on Vertical Electronic Transitions by the Polarizable Continuum Model. *J. Chem. Phys.* **2000**, *112*, 2427-2435.

- (22) Jagoda-Cwiklik, B.; Slavicek, P.; Cwiklik, L.; Nolting, D.; Winter, B.; Jungwirth, P. Ionization of Imidazole in the Gas Phase, Microhydrated Environments, and in Aqueous Solution. *J. Phys. Chem. A* **2008**, *112*, 3499-3505.
- (23) Schlegel, H. B. Potential-Energy Curves using Unrestricted Moller-Plesset Perturbation Theory with Spin Annihilation. *J. Chem. Phys.* **1986**, *84*, 4530-4534.
- (24) Roca-Sanjuan, D.; Rubio, M.; Merchan, M.; Serrano-Andres, L. Ab Initio Determination of the Ionization Potentials of DNA and RNA Nucleobases. *J. Chem. Phys.* **2006**, *125*, 084302.
- (25) Boese, A. D.; Martin, J. M. L. Development of Density Functionals for Thermochemical Kinetics. *J. Chem. Phys.* **2004**, *121*, 3405-3416.
- (26) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, Jr. J. A.; Vreven, T.; Kudin, K. N.; Burant, J. C. et al. Gaussian03, Gaussian Inc.: Wallingfort, CT, 2003.
- (27) Winter, B.; Faubel, M. Photoemission From Liquid Aqueous Solutions. *Chem. Rev.* **2006**, *106*, 1176-1211.
- (28) Seidel, R.; Thurmer, S.; Winter, B. Photoelectron Spectroscopy Meets Aqueous Solution: Studies from a Vacuum Liquid Microjet. *J. Phys. Chem. Lett.* **2011**, *2*, 633-641.
- (29) Lide, D. R. *CRC Handbook of Chemistry and Physics*. Taylor & Francis: New York, 2005.
- (30) Pluharova, E.; Schroeder, C.; Seidel, R.; Bradforth, S. E.; Winter, B.; Faubel, M.; Slavicek, P.; Jungwirth, P. Unexpectedly Small Effect of the DNA Environment on Vertical Ionization Energies of Aqueous Nucleobases. *J. Phys. Chem. Lett.* **2013**, *4*, 3766-3769.
- (31) Papadantonakis, G. A.; Tranter, R.; Brezinsky, K.; Yang, Y. N.; van Breemen, R. B.; LeBreton, P. R. Low-eEnergy, Low-Yield Photoionization, and Production of 8-oxo-2'-deoxyguanosine and Guanine from 2'-deoxyguanosine. *J. Phys. Chem. B* **2002**, *106*, 7704-7712.

(32) Pluharova, E.; Oncak, M.; Seidel, R.; Schroeder, C.; Schroeder, W.; Winter, B.; Bradforth, S. E.; Jungwirth, P.; Slavicek, P. Transforming Anion Instability into Stability: Contrasting Photoionization of Three Protonation Forms of the Phosphate Ion upon Moving into Water. *J. Phys. Chem. B* **2012**, *116*, 13254-13264.

(33) Seidel, R.; Faubel, M.; Winter, B.; Blumberger, J. Single-Ion Reorganization Free Energy of Aqueous  $\text{Ru}(\text{bpy})_3^{2+/3+}$  and  $\text{Ru}(\text{H}_2\text{O})_6^{2+/3+}$  from Photoemission Spectroscopy and Density Functional Molecular Dynamics Simulation. *J. Am. Chem. Soc.* **2009**, *131*, 16127-16137.

TOC GRAPHIC

